Exome sequencing and functional validation in zebrafish identify novel genes for dystroglycanopathies

> M. Chiara Manzini PhD Boston Children's Hospital Manton Center for Orphan Disease Research Harvard Medical School

Spring 2013: Dept. of Pharmacology and Physiology The George Washington University Medical Center

Dystroglycanopathies affect muscle, eye and brain development

- Core features:
- Congenital muscular dystrophy
- Brain malformations: hydrocephalus, gyral defects
- Ocular malformations in the retina, lens and cornea
- Dystroglycan is a widely expressed transmembrane glycoprotein
- α-dystroglycan (α-DG) interacts with extracellular proteins like laminin via its glycosylated moieties





Dystroglycanopathies are extremely genetically and clinically heterogeneous

- Mutations in 10 genes cause dystroglycanopathies and all affect α-DG glycosylation
- Some genes are glycosyltransferases, others are suspected to have a role in glycosylation
- α-DG glycosylation is essential for it function, e.g. laminin binding
- Mutations in these genes explain only a portion of the cases



Mutations in the same gene cause a spectrum of phenotypes affecting the brain-eye-muscle or only the muscle

Brain phenotypes range greatly in severity



Homozygosity mapping provides linkage information at the single family level



Identifying patients with rare diseases requires a large network of collaborators



Many of our collaborators are based in countries with high rate of consanguineous marriages: consanguinity yields more informative families

Homozygosity mapping in a dystroglycanopathy family

- 1st cousin union affected with Walker Warburg Syndrome
- 9.6% homozygosity 2480 genes



Homozygosity mapping and exome sequencing identify a new disease gene

- Exome sequencing yields 30-50,000 variants per individual
- A need for an appropriate filtering strategy
 - In region of homozygosity: focus on the 2480 genes
 - Rare or novel
 - Pathogenic: missense, stop, splicing, frameshift
- This approach reduced the number of candidate genes from 2480 to 6!
- A stop codon in the glycosyltransferase GTDC2 appeared to be the best candidate (Manzini et al, 2012)



The zebrafish larva as a functional model for variant validation

The zebrafish larva has prominent eyes and muscle, which develop over a few days





gtdc2 loss of function severely affects development

- Morphants for gtdc2 show severe developmental defects from 1dpf
 - Stunted growth
 - Disorganized muscle fibers
 - Reduced mobility
 - Stunted head and eye development



Survival reduced to 50% by 3 dpf

gtdc2 loss of function severely affects development

- 3dpf morphant larvae are short, with a bent tail and display impaired mobility
- Normal ventral retinal fusion is prevented





gtdc2 morphants also show severe hydrocephalus



Human GTDC2 mRNA rescues the morphant phenotype





Non-consanguineous families can also be easily studied

- Unrelated parents: two children and one fetus affected with WWS
- Only one gene with compound heterozygous mutations shared by all affected children: B3GALNT2
- Three additional families with alleles in the same gene



B3GALNT2 is another new dystroglycanopathy gene

- B3GALNT2 is another glycosyltransferase to add to the puzzle
- Zebrafish MO knockdown analysis is consistent with the findings from other dystroglycanopathies





Conclusions: Combination of human genetics and zebrafish validation to identify novel disease genes

Homozygosity mapping and WES in consanguineous families can identify disease-causing mutations in rare, severe disorders

WES is effective even in non-consanguineous unions with multiple affected children

The zebrafish embryo is a rapid validation model for candidate mutations: muscle, eye and brain phenotypes are comparable to the ones observed in human patients

GTDC2 and B3GALNT2 are novel dystroglycanopathy genes identified using this approach

Multiple new candidates can be tested in this fashion

Chris Walsh

Muna Al Saffar Brenda Barry Jen Partlow

Whole-exome

Sean Hill Tim Yu

Danielle Gleason Jackie Rodriguez

Erin Heinzen (Duke) David Goldstein (Duke) Stacy Gabriel (Broad)

WWS-zebrafish

Vandana Gupta Alan Beggs Tom Maynard (GWU)

Richard Hawkes (U. Calgary) Masahito Hibi (U. Nagoya)

ASD/ID

Dilenny Gonzales David Tischfield Deniz Atabay Stefania DiCostanzo Vanessa Mitsialis (Harvard) Antonella Cinquino (La Sapienza, Rome) Dimira Tambunan (Northeastern)

Ranad Shaheen (King Faisal) Diya Sabbagh (King Faisal) Fowzan AlKuraya (King Faisal) Mustafa Salih (King Saud)

Funding:

Current: ASD/ID - NICHD/NIH K99, WWS - Manton Center for Orphan Diseases

Past: WWS - MDA, Hearst Fund, ASD/ID - Centro per la Comunicazione e Ricerca (Italy)

Our clinical team



CHB Team: Ganesh Mochida, Ramzi Nasir, Muna Al Saffar, Jen Partlow, Wen-Hann Tan

GTDC2 is expressed in the brain and muscle during human development

- high brain and muscle expression in fetal and adult tissues
- highly expressed in the heart and kidneys, which are also affected in the patients



Gtdc2 is expressed in the brain and eye during mouse development

- high brain expression during embryonic development
- expressed in the eye, in the retina, lens and cornea
- cortical expression is highest during proliferation and migration



gtdc2 mRNA is highly expressed in the brain, eye and muscle

- gtdc2 is expressed throughout development and mostly in the brain, eye, and somite boundaries at 1dpf
- at 3dpf highest expression is in the brain, eye and ear





Three independent alleles in *GTDC2* in our patient cohort



gtdc2 loss of function disrupts organization of the retina and the muscle

- Retinal lamination defects are observed in *gtdc2* morphant eyes
- Both dystrophin and glycosylated dystroglycan are greatly reduced at the myosepta



Neuronal migration and differentiation are severely disrupted











Neuronal migration

Cortical lamination





Disrupted basal lamina, misoriented radial flia

Cortical dysplasia, overmigration



(Manzini & Walsh, 2011)

Post-doctoral positions available

Manzini lab

The George Washington University Medical Center Department of Pharmacology and Physiology Department of Integrative Systems Biology The Institute of Neuroscience



Position is fully funded for 2 years starting summer of 2013

Topics:

- The ECM in neuromuscular disorders
- Role of endosomal trafficking in cognitive development

Email CV and reference information to: chiara.manzini@gmail.com



giovanni ARMENISE HARVARD foundation

<u>Giovanni-Armenise Harvard Summer Fellowship for Italian</u> <u>Undergraduate Students</u>

- For outstanding students in biomedical advanced degrees (1st, 2nd year "specialistica" or 4th-6th year Medicine)
- Summer fellowship to conduct research in a Harvard Medical School lab for 8-10 weeks (2000 Euro for 2012)
- For application information: <u>www.armeniseharvard.org</u>
- Application deadline before Christmas
- ✓ Look us up on Facebook!

How do you approach a heterogeneous disorder?

- Collect a large cohort of patients with accurate clinical description
- Study the frequency and distribution of mutations in known genes to provide genetic testing guidelines
- Study families with no mutations in known genes to identify novel disease causes

Summary: WES identifies GTDC2 as a new WWS gene

- WES can rapidly provide a molecular diagnosis for genetic disorders
- Several patients in our cohort had mutations in known disease genes changing the diagnosis or expanding the phenotypic spectrum
- We identified GTDC2 as a novel WWS gene
- In consanguineous families an average of 5 variants per family is identified
- A validation plan must be in place to analyze the function of these variants